

Blocking Protocols

These protocols have been specifically developed for use with arrays printed on SCHOTT Nexterion™ A⁺ Slides (aminosilane coated). This protocol should not be used with any other slide type.

Microarray slides printed by the Ramaciotti Centre have been baked at 80°C for 2 hours.

Prior to use store slides in clean, dark, dry conditions. Slides are stable for 3 months prior to carrying out the blocking procedure.

Carry out the following procedure immediately prior to hybridisation i.e. on the same day. Never allow slides to dry between steps, immediately transfer them directly from one solution to the next.

Protocol from SCHOTT

1. 1 x 10 to 20 sec in 0.1 % SDS at room temperature.
2. 1 x 10 to 20 sec in dIH₂O at room temperature.
3. 1 x 3 min in dIH₂O at 95°C.
Important: only use this step when PCR-products were spotted e.g. custom array.
4. 1 x 45 min in Pre-Hybridisation Buffer (3 -5xSSC, containing 0.1% SDS and 0.1 mg/ml BSA or 25 ml Nexterion Hyb⁺ 25 ml dIH₂O + 500 mg BSA, volume for 5 slides) at 42°C
5. 1 x 10 to 20 sec in dIH₂O at room temperature.
6. Dry slides immediately.

Alternative Protocol

1. Place the slides in a 0.1% SDS solution in ddH₂O, at 95 °C for 1 minute with constant stirring. (Heat up SDS in microwave until boiling, turn off and place slides in solution for 1 min)
2. Wash in 5 % EtOH solution in ddH₂O for 1 minute at room temp with constant stirring (use 95% EtOH to make up the 5% solution).
3. Wash in double distilled water, or MQ water at room temperature for 1 min with constant stirring.
4. Dry slides by centrifugation at 800 rpm for 8 min.
5. Proceed directly to hybridisation.