

RNA Precipitation Procedure

1. Add 1/10 volume 3M NaOAc, pH 5.2, 2.5 volumes ethanol, and 0.5 μ L of glycogen (5mg/mL)* to your RNA.
 2. Mix and incubate at -80°C for at least 1 hour.
 3. Centrifuge at $\geq 12,000 \times g$ in a microcentrifuge for 20 minutes at 4°C.
 4. Wash pellet twice with 80% ethanol.
 5. Air dry pellet. Check for dryness before proceeding.
 6. Resuspend the pellet in nuclease free H₂O. The appropriate volume for resuspension depends on the expected yield and the amount of RNA required for the labelling. Please read the sample submission form in order to determine the appropriate resuspension volume.
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DNA Cleanup

If a genomic DNA preparation is suspected to contain inhibitors, the following cleanup procedure can be used:

1. Add 0.5 volumes of 7.5 M NH₄OAc, 2.5 volumes of absolute ethanol (stored at -20°C), and 0.5 μ L of glycogen* (5mg/mL) to 250ng genomic DNA.
2. Vortex and incubate at -20°C for 1 hour.
3. Centrifuge at 12,000 $\times g$ in a microcentrifuge at room temperature for 20 minutes.
4. Remove supernatant and wash pellet with 0.5mL of 80% ethanol.
5. Centrifuge at 12,000 $\times g$ at room temperature for 5 minutes.
6. Remove the 80% ethanol and repeat the 80% ethanol wash one more time.
7. Re-suspend the pellet in reduced EDTA TE buffer (10mM Tris, pH 8.0, 0.1mM EDTA, pH 8.0). Please read the sample submission form in order to determine the appropriate resuspension volume.

*Addition of Carrier to Ethanol Precipitations

Adding carrier material has been shown to improve the RNA/DNA yield of precipitation reactions. Addition of 0.5 to 1 μ L of glycogen (5 mg/mL) to nucleic acid precipitations aids in visualisation of the pellet and may increase recovery. The glycogen does not appear to affect the outcome of subsequent steps in this protocol.